



Effect of Henna Addition on the Surface Roughness and Hardness of Polymethylmethacrylate Denture Base Material: An *in vitro* Study

¹Amal Nawasrah, ²Mohammed M Gad, ³Mai El Zayat

ABSTRACT

Aim: This study aimed to evaluate the effect of the addition of various henna—which can have antifungal properties—on the surface roughness and hardness of polymethylmethacrylate (PMMA) denture base material.

Materials and methods: A total of 99 rectangular-shaped ($10 \times 20 \times 3 \text{ mm}^3$) specimens were prepared from heat-cured acrylic resin and divided into one control group without the addition of henna and five test groups, which were prepared by adding *Yamani henna* powder to polymer at concentrations of 1, 2.5, 5, 7.5, and 10 wt%. The polymer was added to the monomer, mixed, packed, and processed using the conventional water bath method. After processing, specimens were finished and polished, then kept in distilled water for 48 ± 2 hours. A profilometer and Vickers hardness tester were used to measure surface roughness and hardness respectively. Statistical data analysis was conducted via Statistical Package for the Social Sciences (SPSS) version 20.0 (IBM, USA). The independent sample t-test was used and $p \leq 0.05$ was considered statistically significant.

Results: The addition of henna at varying concentrations significantly increased the surface roughness values ($p \leq 0.01$) while decreasing hardness ($p \leq 0.0001$). The most favorable addition value was 1% henna between all henna groups.

Conclusion: The addition of henna to the acrylic resin may negatively affect the surface properties of PMMA acrylic denture base.

Clinical significance: Antimicrobial denture with minimum deterioration effects on its physical properties could be achieved with henna addition to denture base material in low concentration. However, 1% henna showed the best results between the henna groups as regards roughness and hardness values.

Keywords: Hardness, Henna, Physical properties, Polymethylmethacrylate, Surface roughness.

How to cite this article: Nawasrah A, Gad MM, El Zayat M. Effect of Henna Addition on the Surface Roughness and Hardness of Polymethylmethacrylate Denture Base Material: An *in vitro* Study. J Contemp Dent Pract 2018;19(6):732-738.

Source of support: Nil

Conflict of interest: None

INTRODUCTION

Denture stomatitis (DS) is an inflammation of the mucosa under the complete denture.¹ A symptomless disease,² its over- or underdiagnosis will lead to problems, such as errors in treatment and abuse of antifungal agents, leading to antimicrobial resistance.³ Instances of DS become more common with increasing age, especially in medicocompromised patients.⁴ *Candida albicans* is a common organism in the oral cavity;⁵ known as a pathogenic species, it may cause not only mucosal but also deep-tissue infections that can reach the bloodstream.⁶ *Candida albicans* is able to adhere to the surface of PMMA denture base material. The presence of plaque and poor oral hygiene will render the organism virulent,⁵ leading to the development of DS.⁷ Its pathogenicity has increased due to excessive use of antibiotics and immunosuppressors.⁸

Treatment of DS includes adjusting the existing denture and following oral hygiene instructions. The attainment of an antimicrobial denture is the aim of many researchers and dentists, who have attempted to make certain modifications to PMMA, such as treating the denture surface⁶ or coating it with hydrophilic material⁹ to reduce the adherence of microbial agents.¹⁰ The treatment of PMMA resins has been effected through various mechanisms, including the addition of antimicrobial

¹⁻³Department of Substitutive Dental Sciences, College of Dentistry, Imam Abdulrahman Bin Faisal University, Dammam Kingdom of Saudi Arabia

Corresponding Author: Mohammed M Gad, Department of Substitutive Dental Sciences, College of Dentistry, Imam Abdulrahman Bin Faisal University, Dammam, Kingdom of Saudi Arabia, Phone: +966592502080, e-mail: dr.gad@hotmail.com

polymer (10% poly (2-tert-butylaminoethyl) methacrylate, (PTBAEMA)) to PMMA to reduce the adhesion and growth of *C. albicans* on the denture bases.¹¹⁻¹³

Among the antimicrobial agents, henna was added to the denture base as an antifungal agent to prevent DS.¹² Henna is a naturally occurring plant that is mainly used for coloring the hair and hands.¹³ In a recent review of the pharmacological effect of henna, it was found that it has anti-inflammatory, antimicrobial, and antioxidant properties.¹⁴⁻¹⁶ In addition, henna has burn wound healing properties¹⁷ and even antitumor effects.¹⁸

The characteristics of surface roughness affect the adhesion and initial biofilm formation of *C. albicans* on acrylic surfaces, most likely because PMMA provides a greater surface area and protected sites for colonization.¹⁹ Some studies have shown a direct correlation between surface roughness and *C. albicans*; both plaque accumulation and said organism's adhesion were enhanced when the surface roughness increased.²⁰

Hardness is defined as the resistance of a material to permanent surface indentation or penetration; it is considered an indicator of the denture base material's abrasiveness. The surface properties of PMMA denture base material can be affected by hardness, which is characteristic of the ease of finishing the material as it is resistant to in-service scratching during cleansing.²¹

Although the addition of henna to PMMA has antimicrobial advantages, the surface properties of PMMA denture base material after such addition have not been investigated. Therefore, the aim of this study was to evaluate the effect of henna incorporation in PMMA on the latter's surface roughness and hardness properties. The hypothesis is that the addition of henna to a PMMA denture base will affect the latter's surface properties.

MATERIALS AND METHODS

This study involved an *in vitro* investigation of 99 specimens ($n=9$) of rectangular-shaped ($10 \times 20 \times 3 \text{ mm}^3$) acrylic plates, prepared using heat-cured acrylic resin

(Major base 20, Major Prodotti Dentari SPA, Italy) and divided into two main groups with different ratio calculations, each group will be divided into six subgroups ($n=9$), group I specimens' powder = AR + henna. Group II specimens' powder = AR only + henna as additive to which were added Yamani henna powder (Harazi, Sanaa, Yeman) at concentrations of 0, 1, 2.5, 5, 7.5, and 10 wt%.

An electronic balance (Denver instrument GmbH, Goettingen, Germany) was used to weigh the henna, which was then added to the PMMA powder inside bottles and shaken manually for around three minutes each.

The first group of 54 specimens was distributed into six subgroups ($n=9$) (Table 1). For the first group, the powder was the sum of both the PMMA and henna powder with a powder-liquid ratio of 1:1.²² Meanwhile in the second group, the remaining 45 specimens were distributed into five subgroups; the control group was the same for both groups ($n=9$). In the second group, henna was added to the PMMA powder; therefore, the powder consisted completely of PMMA-liquid (1:1). Both groups were used to test the roughness and hardness, using a profilometer and hardness tester respectively.

A silicone mold with internal dimensions of $10 \times 20 \times 3 \text{ mm}^3$ was used to fabricate 99 wax specimens (Cavex setup wax, Cavex, the Netherlands). The wax specimens were invested in dental stone (Talladium dental stone, USA). After dewaxing the randomly allocated flasks (61B Two Flask Compress, Handler Manufacturing, US), the mold spaces were cleaned and a thin separating media was applied (Isolmajor, Major Prodotti Dentari SPA, Italy). The PMMA-henna composites were mixed with the monomer until a doughy consistency was attained, before being packed into the mold. The specimens were placed in an automatic polymerization water tank (KaVo Elekto Technisches WERK GmbH D-88299 Leutkirch im Allgau, TYP 5506, Germany). Temperature was maintained at 73°C for 90 minutes, before being increased to

Table 1: Ratio of henna to acrylic resin powder in both groups

Group	Subgroups	Code	Polymer conc.(gm)	Henna conc.(gm)	Total (gm)	#
Control	0%		100	0	100	9
Group I	1%		99	1	100	9
	2.5%		97.5	2.5	100	9
	5%		95	5	100	9
	7.5%		92.5	7.5	100	9
	10%		90	10	100	9
Group II	1%		100	1	101	9
	2.5%		100	2.5	102.5	9
	5%		100	5	105	9
	7.5%		100	7.5	107.5	9
	10%		100	10	110	9

100°C for 30 minutes. After processing, flasks were left aside for bench cooling for at least 5 hours before deflasking. The deflasked specimens were finished and polished using the Buehler MetaServ 250 Polisher and Grinder. Specimens were polished with a polishing cloth and a Masterprep Polishing Solution (0.05 micron) at speed using water as a cooling agent then stored in distilled water at 37°C for 48 ± 2 hours before testing.²³

TESTING PROCEDURES

The testing of the surface roughness was performed using the appropriate tester (Contour GT, Bruker Nano GmbH Schwarzschildstrasse 12, 12489 Berlin, Germany). A linear variable differential transformer was deployed to measure the surface morphology, while the numerical values of the surface profile were calculated in a computer to obtain the Ra (the roughness of a surface, which is the mean value of all absolute distances of the roughness profiles from the mean line within the measuring distance using the noncontact lens during tracing).

The hardness test was conducted using the corresponding tester (Wilson Hardness, ITW Test and Measurement GmbH, Shanghai, China) equipped with a Vickers diamond. Eight indentations were made per specimen through the application of a 25-gf load for 30 seconds. To attain the final hardness values, the average of the eight indentations was calculated for each specimen.²³

Statistical Analysis

Statistical data analysis was performed using SPSS version 20.0 (IBM, USA). The results of the surface roughness and hardness tests were shown as arithmetic mean and standard deviation (SD). In comparisons with the control group, an independent sample t-test used. A p-value ≤ 0.05 was considered a statistically significant result.

RESULTS

The mean values and SD of the Ra are summarized in Table 2; it can be observed that the henna powder significantly increased in Ra value for all concentrations in both groups (p-value ≤ 0.01), in comparison with the control

group. The addition of 1% powder returned the lowest Ra value (0.0501 ± 0.0052) under the micrometer, while the 10% addition resulted in the highest value ($0.316.9 \pm 0.0351$).

Among the groups, there were significant differences between those with 1 and 7.5%, while there were no significant differences between other groups. Figures 1 and 2 show the average surface roughness values using parameters with two colors (red and blue). Red denotes the peak's height, while blue reflects the valley's depth, or deviation, from a mean line. The graduated colors between red and blue (the readings for the top and bottom) in this parameter represent the overall surface roughness (Figs 1 and 2).

Table 3 shows the mean values and SD of the hardness test. Statistical analysis revealed that the hardness reduced significantly (p-value ≤ 0.0001) in comparison with the control group, as well as between groups. As the henna concentration increased, the hardness decreased.

DISCUSSION

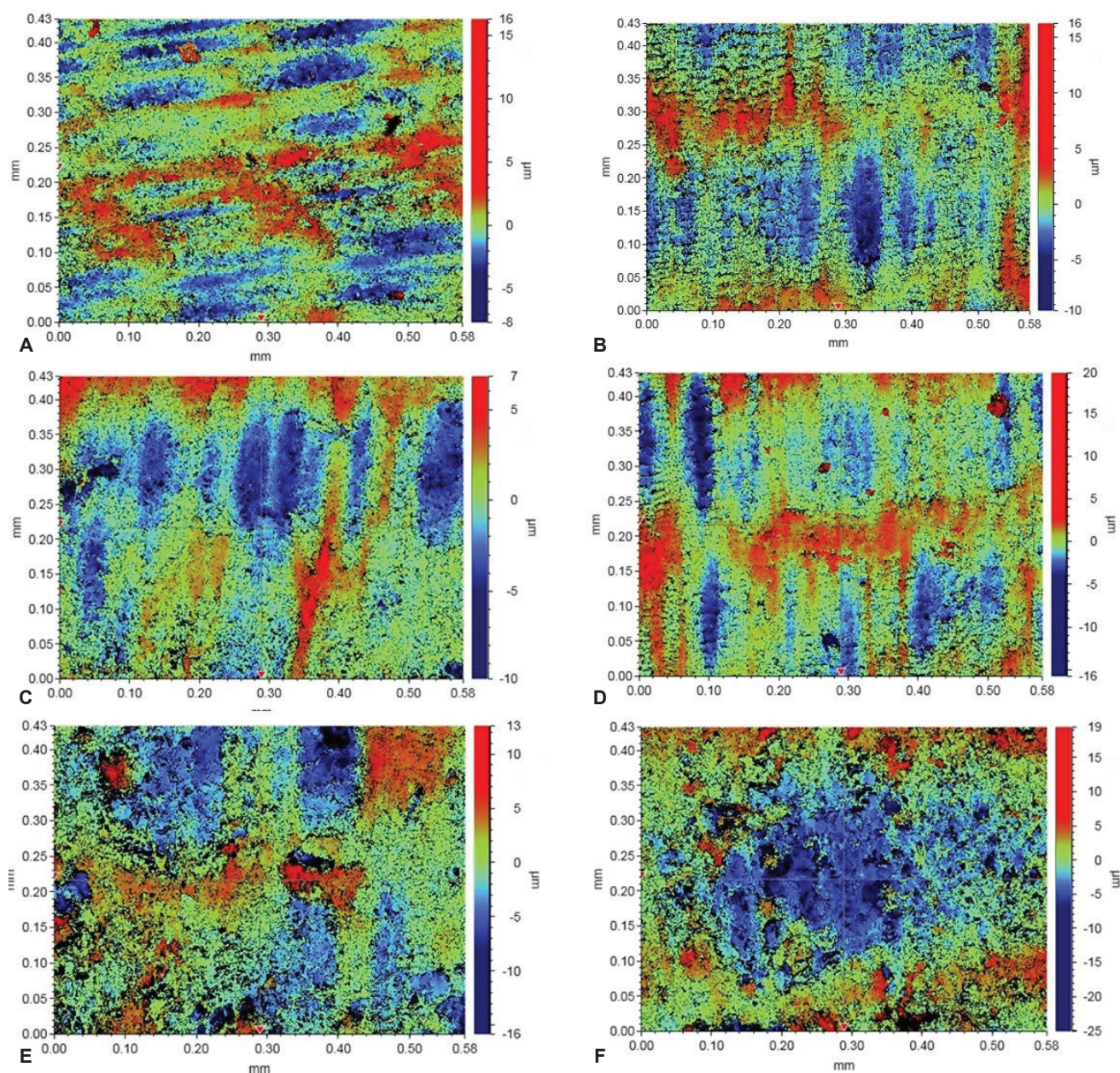
With the aim of improving the oral health of denture wearers, many studies have suggested the addition of antifungal agents to PMMA acrylic resin, which may represent a possible alternative method of controlling oral infections. Although henna powder has an antifungal effect when added to the PMMA denture base material, it may alter the latter's surface characteristics. Accordingly, this *in vitro* study was carried out to evaluate the effect of henna on surface properties, with the results indicating that said properties affected especially with high henna concentrations. Therefore, the hypothesis was accepted.

The surface properties of denture base material are important as they affect the oral health of tissues in direct contact with the dentures. Since the initial adhesion of microorganisms is directly influenced by the surface roughness,²⁴ the latter can increase or decrease microbial adhesion, colonization, and biofilm maturation.^{25,26} It has been reported that resins with high Ra values displayed higher numbers of *C. albicans* in comparison with highly polished smooth surfaces, possibly due to the increased hills and peaks of the rougher surfaces, which provide a wide surface area and are more accessible active sites

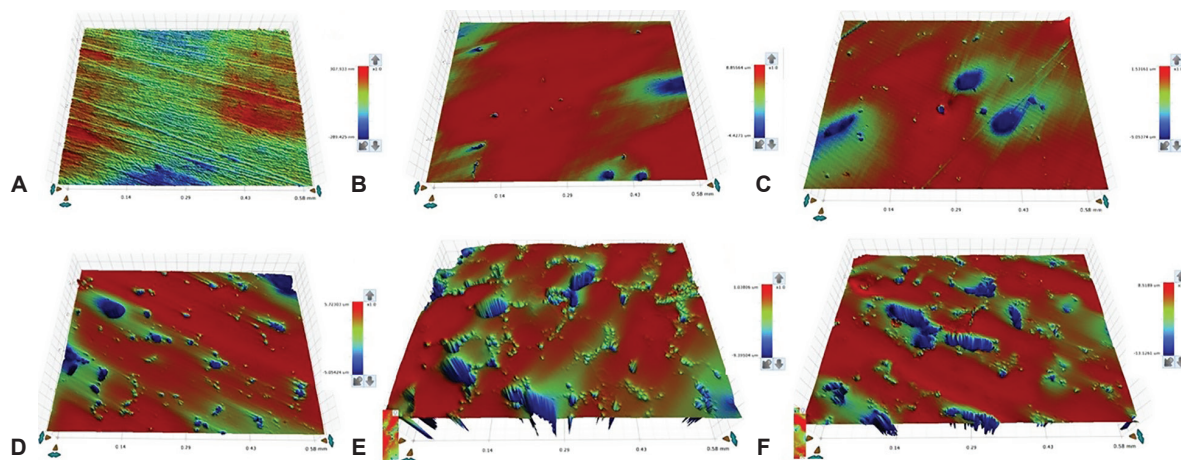
Table 2: Main values and SD of surface roughness (μm) results

	Group I		Group II		Mean comparison b/w groups
	Mean (SD)	p-value	Mean (SD)	p-value	
0%	0.01738 \pm 0.0016		0.01738 \pm 0.0016		
1%	0.0501 \pm 0.0052	0.0001*	0.0398 \pm 0.0021	0.0001*	0.0001*
2.5%	0.0556 \pm 0.0069	0.0001*	0.0552 \pm 0.0059	0.0001*	0.44
5%	0.1568 \pm 0.0132	0.0001*	0.1499 \pm 0.0139	0.0001*	0.089
7.5%	0.2436 \pm 0.0343	0.0001*	0.2778 \pm 0.0337	0.0001*	0.0058*
10%	0.316.9 \pm 0.0351	0.0001*	0.3222 \pm 0.0429	0.0001*	0.36

*Statistically significant



Figs 1A to F: Two-dimensional representative surface roughness images of tested specimens according to henna concentrations: (A) 0% henna, (B) 1% henna, (C) 2.5% henna, (D) 5% henna, (E) 7.5% henna, and (F) 10% henna



Figs 2A to F: Three-dimensional representative surface roughness images of tested specimens according to henna concentrations: (A) 0% henna, (B) 1% henna, (C) 2.5% henna, (D) 5% henna, (E) 7.5% henna, and (F) 10% henna

Table 3: Main values and SDs of hardness (kg/mm²) results

	Group I		Group II		Mean comparison b/w groups
	Mean ± SD	p-value	Mean ± SD	p-value	
0%	30.1 ± 0.58		30.1 ± 0.58		
1%	29.5 ± 0.77	0.013	30.5 ± 0.39	0.003	0.0001
2.5%	29.1 ± 0.62	0.0001	29.6 ± 0.399	0.0001	0.0095
5%	24.1 ± 0.36	0.0001	23.6 ± 0.38	0.0001	0.00093
7.5%	19.9 ± 0.39	0.0001	18.8 ± 0.33	0.0001	0.0001
10%	15.4 ± 0.2	0.0001	15.7 ± 0.31	0.0001	0.0026

All p-values are statistically significant

for microbial adhesion.^{27,28} Lower surface roughness is important to prevent stain catching.²⁹

The results of the current study showed that henna, as an addition to the PMMA denture base material, significantly increased the surface roughness (Ra) of the specimens (p-value ≤ 0.01). The increase in Ra values with high henna concentrations may be attributed to the existing loosely attached clusters of henna particles on the resin surface, which are easily removed using finishing and polishing techniques, leaving voids.

A removable prosthesis with 0.2 µm surface roughness is considered acceptable for clinical use²⁷ and plaque accumulation of more than this value is expected in prosthetics.^{30,31} This increases the microbial adhesion, which is considered critical in impacting the value of PMMA roughness.³² Ra values found in this study for henna groups of low concentrations (1, 2.5, 5%) were below this level and considered clinically acceptable. In contrast, with the increased concentrations of 7.5 and 10%, the values are more than the clinically acceptable threshold. Therefore, 5% henna could be considered as the limit, below which microbial adhesion reduction might be anticipated.

Compagnoni et al¹¹ studied the effect of antimicrobial polymer PTBAEMA on the surface roughness and found that the addition of PTBAEMA to a PMMA acrylic denture base increased the Ra values, which is in line with the results of the current study. Meanwhile, Regis et al²¹ showed that the use of antimicrobial monomer methacryloyloxyundecylpyridinium bromide (MUPB) had no effect on the roughness of polished acrylic resin. Another team used alcohol vinegar in the PMMA as a way to reduce the number and adhesion of *C. albicans*; this had no effect on the surface roughness of the PMMA.³³ The differences in results may be due to the nature of the added antimicrobial agents, concentrations, methods of additions, or the PMMA denture base material used. Although clear correlations have been reported between surface roughness and microbial adhesion and biofilm formation,²⁵ one group reported no significant influence of roughness on adherence of *C. albicans*.³⁴ Therefore,

whether surface roughness enhances or reduces microbial adhesion remains a matter for debate.

The hardness test has been considered a simple and useful method to determine the mechanical properties of polymer-based materials due to its high sensitivity to the amount of monomer in dental polymers.³⁵ Hardness is important in preventing abrasion and wearing of the PMMA denture base.²⁸

It was observed that the hardness significantly decreased after the addition of henna at all concentrations. This may imply that the incorporation of henna interferes with the conversion of the tested resin. A possible reason for such an effect is the incomplete polymerization process of PMMA denture base material after the addition of henna, resulting in a large amount of residual monomer. The latter adversely affects the mechanical properties via a plasticizing effect.³⁶ In addition, increased henna concentrations that are above the saturation limit act as impurities within the resin matrix, subsequently weakening the materials and affecting their mechanical properties. As we have seen, increasing the henna concentration decreased the hardness of the PMMA resins. This may be explained by the fact that the maximum saturation of the matrix formation between PMMA and henna occurred at concentrations of 1 and 2.5%.³⁷

With the aim of improving the properties of the PMMA denture base, antimicrobial polymers were added. Regis et al²¹ found that the incorporation of 0.6% antimicrobial MUPB reduced the base's hardness, in agreement with the results of the current study. That said, our results are in opposition to a study in which alumina was added to the PMMA, which had a positive impact on the latter's physical and mechanical properties.³⁰ The increase in hardness may have been due to the characteristics of the alumina particles, which possess strong ionic interatomic bonding, giving rise to their desired material features that are not found in henna.³⁸

Recently, nanotechnological products have been incorporated into PMMA to induce antimicrobial properties; their effects on the mechanical properties have been evaluated.^{39,40} For example, Köroğlu et al³⁹ studied the effect of silver nanoparticle (AgNP) addition on PMMA's mechanical and thermal characteristics, concluding that the incorporation of AgNPs, which are generally used for antimicrobial efficiency, affected the mechanical properties of the PMMA denture base depending on the concentration of nanoparticles. In contrast, Gad et al⁴⁰ reported that zirconia nanoparticles have an antifungal effect without affecting mechanical properties. The differences in results may be due to surface properties of added particles as well as the bond between added particles and PMMA matrix.

In group I in our study, henna was added as a part of the acrylic powder ratio, while in group II, it was an

additive over the proposed powder ratio. This was done to evaluate the effect of the addition method on the properties of the final products. Table 2 shows comparative values between groups I and II and displays a significant difference in Ra values. With low concentrations, values of both groups were close, while group II showed higher values with increased henna concentrations. This confirms that the additive method, with increased concentrations, affects the polymer–monomer ratio, whereas excess henna particles interfere with the polymerization reaction. Table 3 shows comparative values between groups I and II and displays a significant difference in hardness values. The results of both groups were in close conjunction, with a slight increase in values for group I. Based on the results of this study, as regards a comparison between the two groups, group I is more conventional and could be used for addition; however, this is still ambiguous and further investigations are required at low concentrations.

Clinical implications: The production of a denture base with an antimicrobial effect would be a remarkable achievement. At the same time, the physical and mechanical properties should not be unduly altered. Based on the results of this study, the addition of henna at a high concentration adversely affects the base's physical properties, while all values obtained for tested properties were higher than what is required in accordance with the ISO: 1567 standard requirements. However, low concentrations were closely aligned with ISO: 1567 No. 12. The addition of henna to the PMMA denture base material should be done with caution, as it may adversely affect the latter's physical properties. It is necessary to study the minimum inhibitory concentration of henna as an antifungal agent. There are various schools of thought as regards the processing of the PMMA denture base material; one is to make the denture on two layers by adding just enough henna to form the intaglio surface of the palatal part of the denture. Another way of thinking is to evaluate the use of microwave processing instead of the conventional hot water bath technique to improve the physical properties of the henna-augmented PMMA denture base material.

LIMITATIONS OF THE STUDY

In clinical terms, the results of this study should be taken with caution, as it was performed *in vitro* where the oral environment is different (lacking dynamic movements, saliva with prospective pH, and the presence of other microorganism); this may affect the denture base surfaces and *C. albicans* adhesion. One limitation was the use of just one type of denture base material (i.e., PMMA), as it is the most commonly used type in this field. In addition, we could not achieve our goal of retesting the specimens after

a certain time had elapsed, to evaluate the effect of aging on the physical properties of PMMA denture base material containing henna. It is essential to conduct further investigations on henna through coating or attempting to enhance the bonding using the resin matrix. In future, it is also warranted to assess the durability through aging procedures that simulate oral conditions.

CONCLUSION

Within the limitations of this study, it could be concluded that the addition of henna to PMMA denture base material significantly increases the latter's surface roughness and decreases its hardness. The use of henna as an additive constitutes no significant difference from considering it as part of the PMMA powder. A henna concentration of 1% showed the best results between the henna groups as regards roughness and hardness values.

ACKNOWLEDGMENTS

The authors would like to acknowledge the Deanship of Scientific Research, University of Dammam for providing a research grant for this study (Grant No. 2014176). They would also like to sincerely thank Miss Joan M Coloma for her assistance in preparing the specimens. Gratitude is also extended to Mr Lindsey Mateo and Mr Soban Khan for their assistance with the mechanical testing and statistical analysis respectively.

REFERENCES

1. Yarborough A, Cooper L, Duqum I, Mendonça T, McGraw K, Stoner L. Evidence regarding the treatment of denture stomatitis. *J Prosthodont* 2016 Jun;25(4):288-301.
2. Walsh T, Riley P, Veitz-Keenan A. Interventions for managing denture stomatitis (Protocol). *Cochrane Database Syst Rev* 2015 Oct;10:CD011920.
3. Gutiérrez O, Juan C, Pérez JL, Oliver A. Lack of association between hypermutation and antibiotic resistance development in *Pseudomonas aeruginosa* isolates from intensive care unit patients. *Antimicrob Agents Chemother* 2004 Sep;48(9):3573-3575.
4. Ishikawa KH, Mayer MP, Miyazima TY, Matsubara VH, Silva EG, Paula CR, Campos TT, Nakamae AE. A multispecies probiotic reduces oral *Candida* colonization in denture wearers. *J Prosthodont* 2015 Apr;24(3):194-199.
5. Sardi JC, Scorzoni L, Bernardi T, Fusco-Almeida AM, Mendes Giannini MJ. *Candida* species: current epidemiology, pathogenicity, biofilm formation, natural antifungal products and new therapeutic options. *J Med Microbiol* 2013 Jan;62(Pt 1):10-24.
6. Filioti J, Spiroglou K, Roilides E. Invasive candidiasis in pediatric intensive care patients: epidemiology, risk factors, management, and outcome. *Intensive Care Med* 2007 Jul;33(7):1272-1283.
7. Paranhos HF, Silva-Lovato CH, Souza RF, Cruz PC, Freitas KM, Peracini A. Effects of mechanical and chemical methods on denture biofilm accumulation. *J Oral Rehabil* 2007 Aug;34(8):606-612.

8. Scully C, el-Kabir M, Samaranayake LP. Candida and oral candidosis: a review. *Crit Rev Oral Biol Med* 1994 May; 5(2):125-157.
9. Yoshijima Y, Murakami K, Kayama S, Liu D, Hirota K, Ichikawa T, Miyake Y. Effect of substrate surface hydrophobicity on the adherence of yeast and hyphal Candida. *Mycoses* 2010 May;53(3):221-226.
10. Gendreau L, Loewy ZG. Epidemiology and etiology of denture stomatitis. *J Prosthodont* 2011 Jun;20(4):251-260.
11. Compagnoni MA, Pero AC, Ramos SM, Marra J, Paleari AG, Rodriguez LS. Antimicrobial activity and surface properties of an acrylic resin containing a biocide polymer. *Gerodontology* 2014 Sep;31(3):220-226.
12. Nawasrah A, AlNimr A, Ali AA. Antifungal effect of henna against *Candida albicans* adhered to acrylic resin as a possible method for prevention of denture stomatitis. *Int J Environ Res Public Health* 2016 May;13(5):E520.
13. Al-Thobity AM, Al-Khalifa KS, Gad MM, Al-Hariri M, Ali AA, Alnassar T. *In vitro* evaluation of the inhibitory activity of thymoquinone in combatting *Candida albicans* in denture stomatitis prevention. *Int J Environ Res Public Health* 2017 Jul;14(7):E743.
14. Abdulmoneim Saadabi MA. Evaluation of Lawsonia inermis Linn. (Sudanese Henna) leaf extracts as an antimicrobial agent. *Res J Biol Sci* 2007 Jan;2(4):419-423.
15. Singla S, Gupta R, Puri A, Bhardawaj VS, Roy S. Comparison of anticandidal activity of Punica granatum (Pomegranate) and Lawsonia inermis (Henna leaves): an *in-vitro* study. *Int J Dent Res* 2013;1(1):8-13.
16. Muhammad HS, Muhammad S. The use of Lawsonia inermis Linn. (henna) in the management of burn wound infections. *Afr J Biotechnol* 2005 Sep;4(9):934-937.
17. Jha RK, Jha PK, Rana SV, Guha SK. Spermicidal action of styrene maleic anhydride polyelectrolyte in combination with magnetic and electrically conductive particles. *Int J Pharmacol* 2009 Jan;5(1):1-12.
18. Lamfon H, Porter SR, McCullough M, Pratten J. Formation of *Candida albicans* biofilms on non-shedding oral surfaces. *Eur J Oral Sci* 2003 Dec;111(6):465-471.
19. Radford DR, Watson TF, Walter JD, Challacombe SJ. The effects of surface machining on heat cured acrylic resin and two soft denture base materials: a scanning electron microscope and confocal microscope evaluation. *J Prosthet Dent* 1997 Aug;78(2):200-208.
20. Pinto Lde R, Acosta EJ, Távora FF, da Silva PM, Porto VC. Effect of repeated cycles of chemical disinfection on the roughness and hardness of hard relined acrylic resins. *Gerodontology* 2010 Jun;27(2):147-153.
21. Regis RR, Zanini AP, Della Vecchia MP, Silva-Lovato CH, Oliveira Paranhos HF, de Souza RF. Physical properties of an acrylic resin after incorporation of an antimicrobial monomer. *J Prosthodont* 2011 Jul;20(5):372-379.
22. Yusuf M, Ahmad A, Shahid M, Khan MI, Khana SA, Manzoor N, Mohammad F. Assessment of colorimetric, antibacterial and antifungal properties of woollen yarn dyed with the extract of the leaves of henna (*Lawsonia inermis*). *J Clean Prod* 2012 May;27:42-50.
23. Kolenbrander PE, Andersen RN, Blehert DS, Eglund PG, Foster JS, Palmer RJ Jr. Communication among oral bacteria. *Microbiol Mol Biol Rev* 2002 Sep;66(3):486-505.
24. Cunha TR, Regis RR, Bonatti MR, de Souza RF. Influence of incorporation of fluoroalkyl methacrylates on roughness and flexural strength of a denture base acrylic resin. *Appl Oral Sci* 2009 Mar-Apr;17(2):103-107.
25. Zamperini CA, Machado AL, Vergani CE, Pavarina AC, Giampaolo ET, da Cruz NC. Adherence in vitro of *Candida albicans* to plasma treated acrylic resin. Effect of plasma parameters, surface roughness and salivary pellicle. *Arch Oral Biol* 2010 Oct;55(10):763-770.
26. Verran J, Maryan CJ. Retention of *Candida albicans* on acrylic resin and silicone of different surface topography. *J Prosthet Dent* 1997 May;77(5):535-539.
27. Radford DR, Sweet SP, Challacombe SJ, Walter JD. Adherence of *Candida albicans* to denture-base materials with different surface finishes. *J Dent* 1998 Sep;26(7):577-583.
28. Consani RL, Folli BL, Nogueira MC, Correr AB, Mesquita MF. Effect of polymerization cycles on gloss, roughness, hardness and impact strength of acrylic resins. *Braz Dent J* 2016 Mar-Apr;27(2):176-180.
29. Arora P, Singh SP, Arora V. Effect of alumina addition on properties of poly-methyl methacrylate-a comprehensive review. *Int J Biotech Trends Technol* 2015 Jan-Mar;5(1):1-7.
30. Bollen CM, Lambrechts P, Quirynen M. Comparison of surface roughness of oral hard materials to the threshold surface roughness for bacterial plaque retention: a review of the literature. *Dent Mater* 1997 Jul;13(4):258-269.
31. Zissis AJ, Polyzois GL, Yannikakis SA, Harrison A. Roughness of denture materials: a comparative study. *Int J Prosthodont* 2000 Mar-Apr;13(2):136-140.
32. Moura JS, da Silva WJ, Pereira T, Del Bel Cury AA, Rodrigues Garcia RC. Influence of acrylic resin polymerization methods and saliva on the adherence of four Candida species. *J Prosthet Dent* 2006 Sep;96(3):205-211.
33. de Castro RD, Mota AC, de Oliveira Lima E, Batista AU, de Araújo Oliveira J, Cavalcanti AL. Use of alcohol vinegar in the inhibition of *Candida spp.* and its effect on the physical properties of acrylic resins. *BMC Oral Health* 2015 Apr;15:52.
34. Azevedo A, Machado AL, Vergani CE, Giampaolo ET, Pavarina AC. Hardness of denture base and hard chair-side relined acrylic resins. *J Appl Oral Sci* 2005 Sep;13(3):291-295.
35. Faltermeier A, Rosentritt M, Müssig D. Acrylic removable appliances: comparative evaluation of different post-polymerization methods. *Am J Orthod Dentofacial Orthop* 2007 Mar;131(3):301.e16-301.e22.
36. Aydogan Ayaz E, Durkan R, Bagis B. The effect of acrylamide incorporation on the thermal and physical properties of denture resins. *J Adv Prosthodont* 2013 May;5(2):110-117.
37. Kati FA, Al-Kaabi AF. Effect of oil paint addition on micro hardness of acrylic ocular prosthesis. *Iraq Dent J* 2016 Aug;38(2): 87-89.
38. Faraji G, Asadi P. Characterization of AZ91/alumina nanocomposite produced by FSP. *Mater Sci Eng A* 2011 Mar;528(6): 2431-2440.
39. Köroğlu A, Ahin O, Kürkçüoğlu I, Dede DO, Özdemir T, Hazer B. Silver nanoparticle incorporation effect on mechanical and thermal properties of denture base acrylic resins. *J Appl Oral Sci* 2016 Nov-Dec;24(6):590-596.
40. Gad MM, Al-Thobity AM, Shahin SY, Alsaqer BT, Ali AA. Inhibitory effect of zirconium oxide nanoparticles on *Candida albicans* adhesion to repaired polymethyl methacrylate denture bases and interim removable prostheses: a new approach for denture stomatitis prevention. *Int J Nanomedicine* 2017 Jul;28(12): 5409-5419.